

**PREMIER Symposium**

Berlin, Germany

March 16<sup>th</sup>, 2018

*u*<sup>b</sup>

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<sup>b</sup>  
**UNIVERSITÄT  
BERN**

# The standardization fallacy

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# IS THERE A REPRODUCIBILITY CRISIS?

A *Nature* survey lifts the lid on  
how researchers view the 'crisis'  
rocking science and what they  
think will help.

BY MONYA BAKER

**52%**  
Yes, a significant  
crisis

**38%**  
Yes, a slight  
crisis

**7%**  
Don't know

**3%**  
No, there is no crisis

**1,576**  
RESEARCHERS SURVEYED

## Things proposed to cause poor reproducibility

- Lack of scientific rigor (risks of bias)
- Too small sample sizes (lack of statistical power)
- “Analytical flexibility” (p-hacking, HARKing, selective reporting)
- Publishing “positive” results only (publication bias)

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- **Rigorous standardization**

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  - Publishing “positive” results only (publication bias)
  - **Rigorous standardization**
- ▶ **Reproducibility is a function of external validity**

# External validity and reproducibility

Birth of *reproducibility* as key principle to establish “matters of fact”



## Open Science in the 17<sup>th</sup> century

Under the eyes of Royal Society members, Robert Hooke replicates an observation reported by a Dutch scientist.

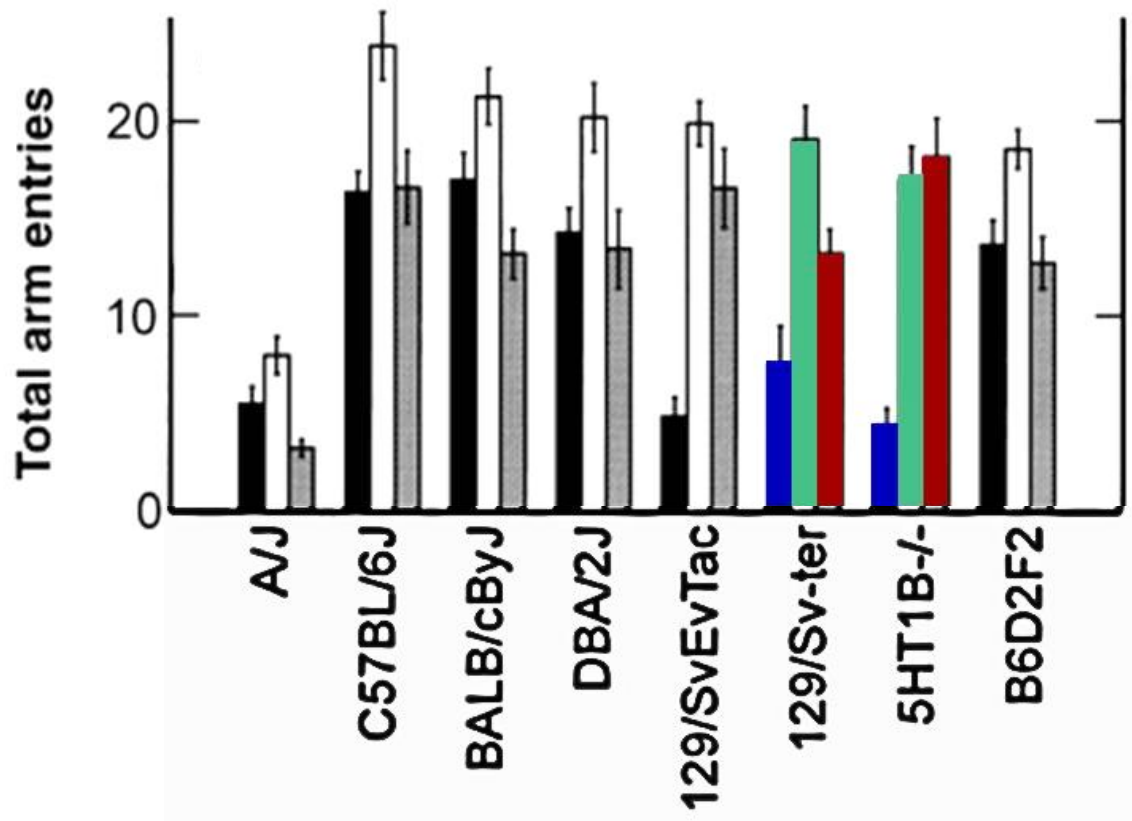
Rita Greer 2007 The Scientists  
Source: **Wikimedia Commons**



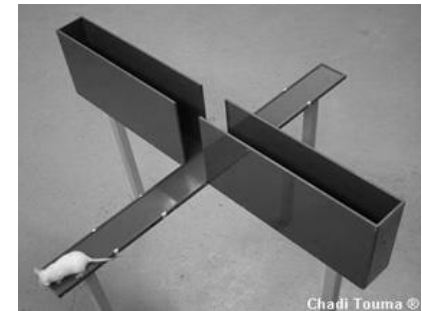


# External validity and reproducibility

Poor reproducibility “despite” rigorous standardization



■ Portland ↗  
■ Edmonton →  
■ Albany ↘





# External validity and reproducibility

## The standardization fallacy

*«Standardization is the attempt to increase reproducibility at the expense of external validity.»*

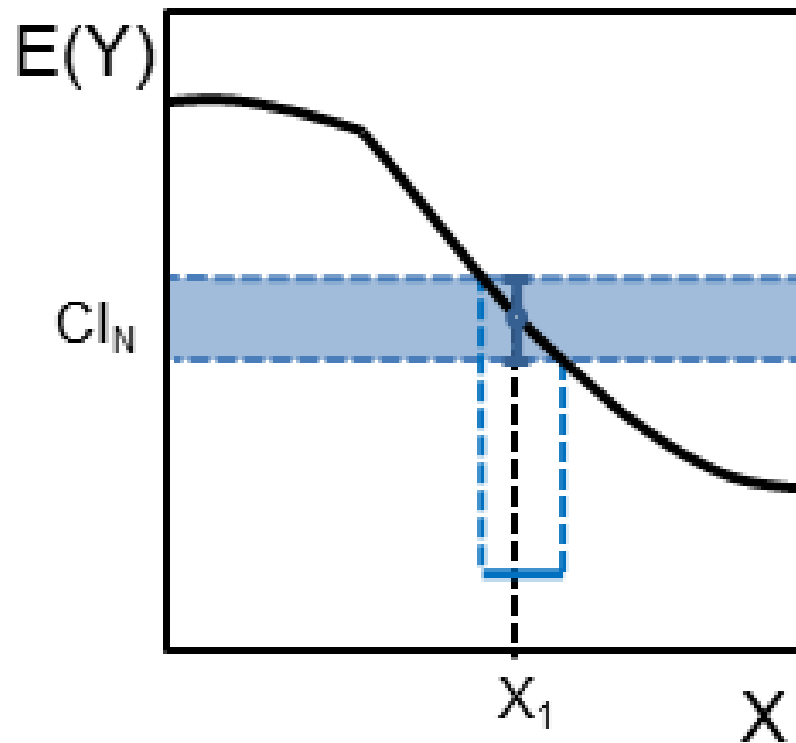
Würbel 2000 **Nature Genetics**

*«A highly standardized experiment supplies direct information only in respect of the narrow range of conditions achieved by standardization. **Standardization, therefore, weakens rather than strengthens our ground for inferring a result, when, as is the case in practice, these conditions are somewhat varied.**»*

Fisher 1935 **The Design of Experiments**

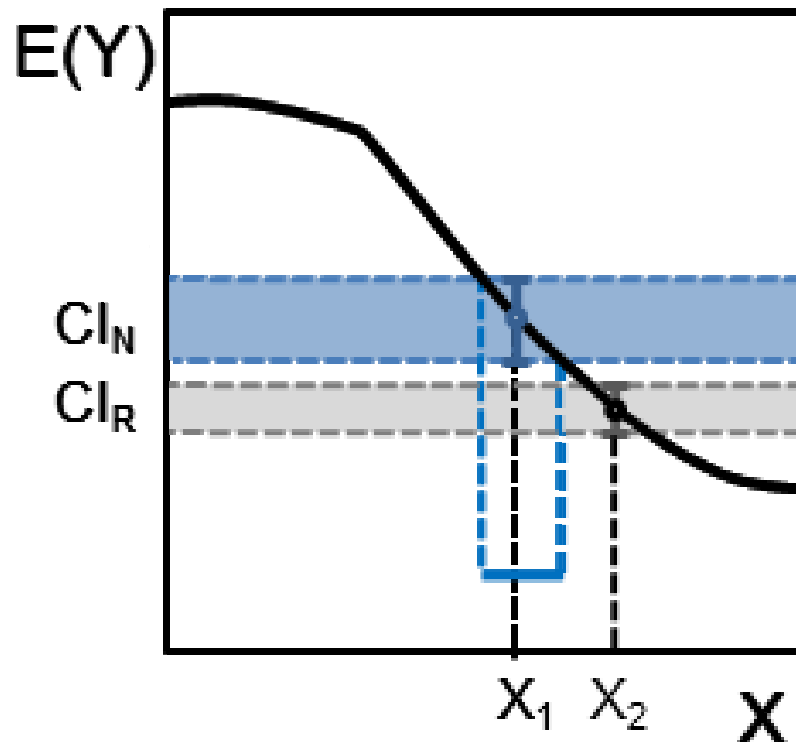
# Standardization and poor reproducibility

## The reproducibility paradox



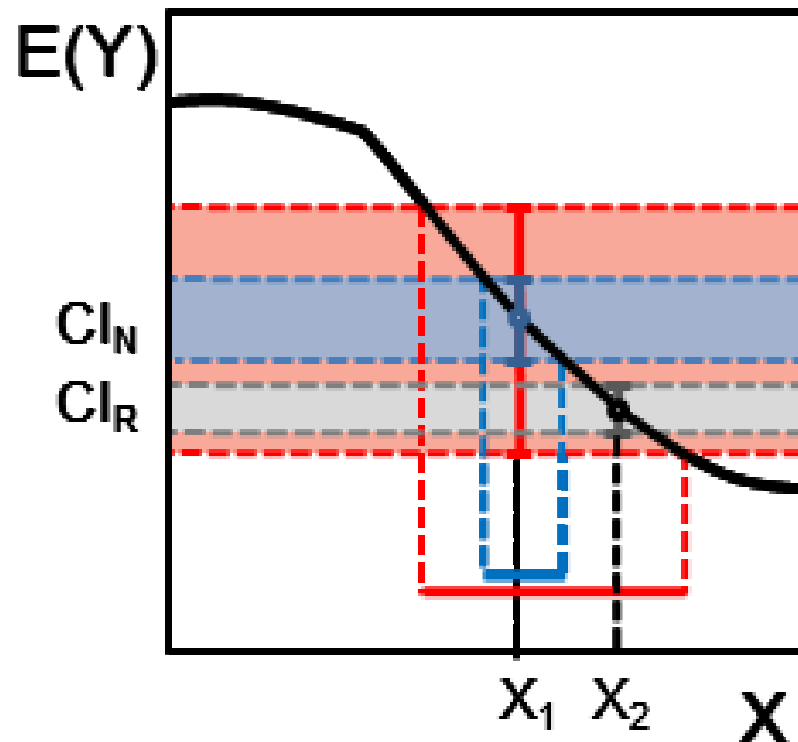
# Standardization and poor reproducibility

## The reproducibility paradox



# Standardization and poor reproducibility

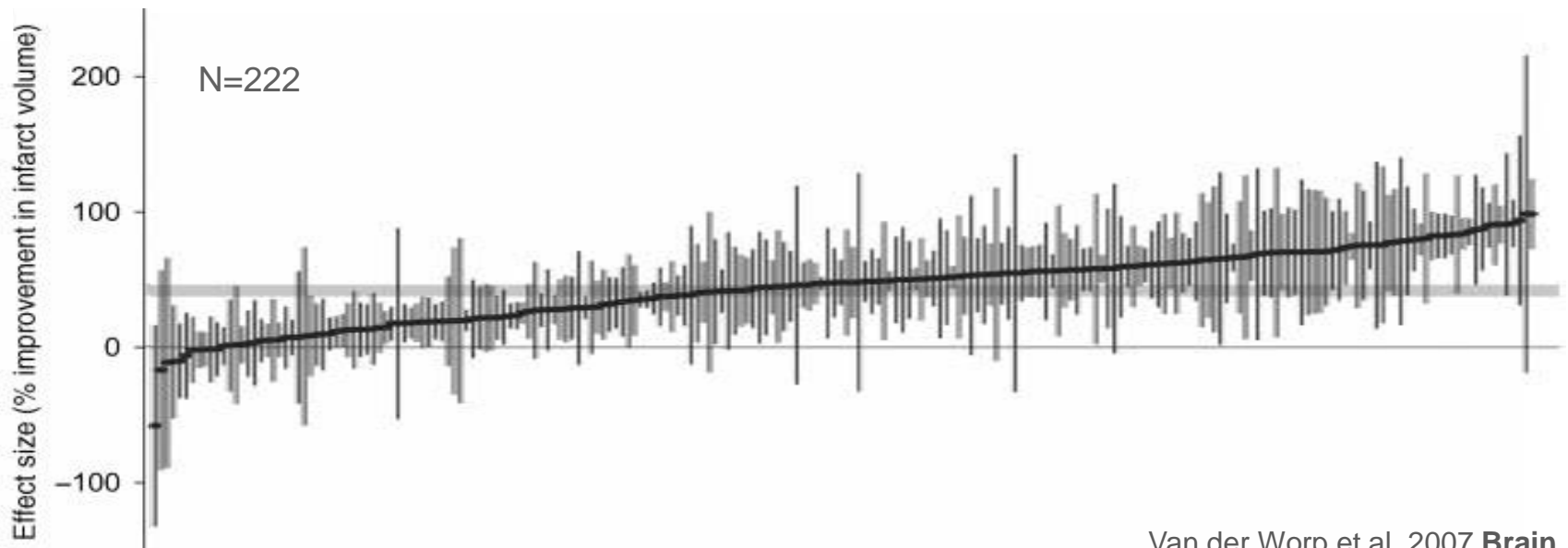
## The reproducibility paradox





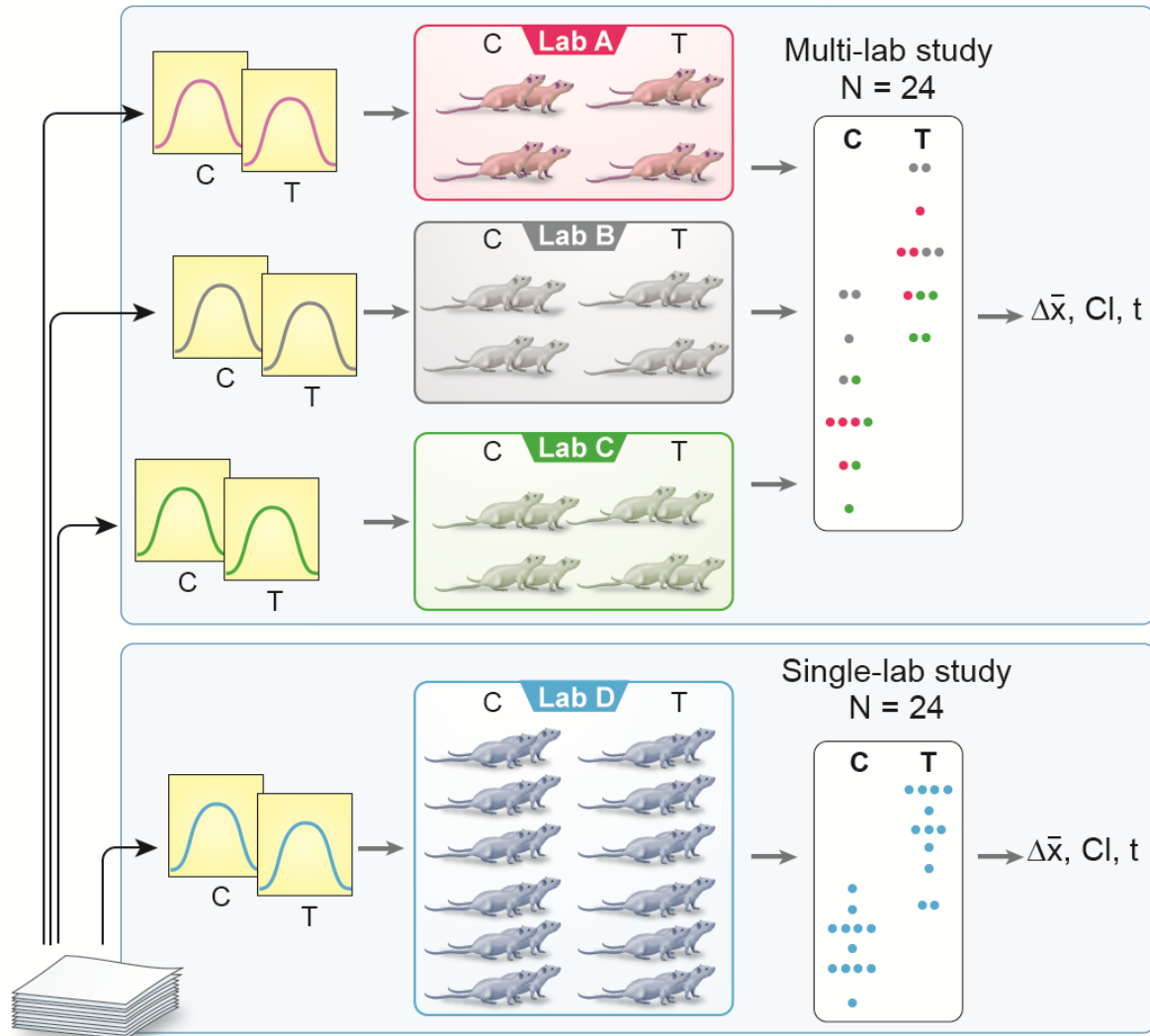
# Simulation of multi-lab studies using real data

## Effect of hypothermia on infarct volume in animal models of stroke



Van der Worp et al. 2007 **Brain**

## Simulation of multi-lab studies using real data

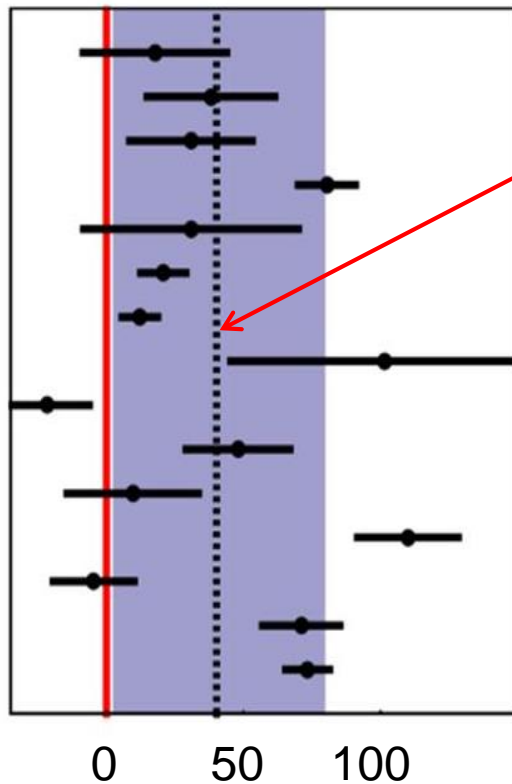


## 50 Studies

# Simulation of single-lab versus multi-lab studies

Effect of hypothermia on infarct volume in rodent models of stroke

1-lab studies

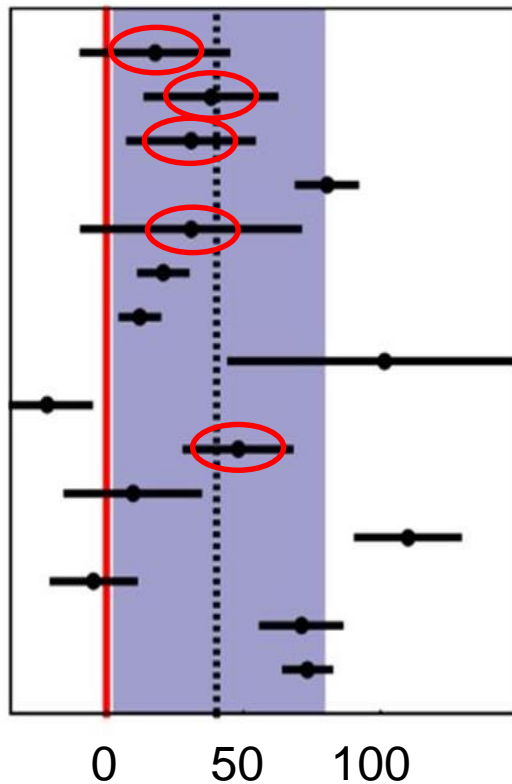


**„true” effect size:**  
summary effect size across all 50 studies

# Simulation of single-lab versus multi-lab studies

Effect of hypothermia on infarct volume in rodent models of stroke

1-lab studies



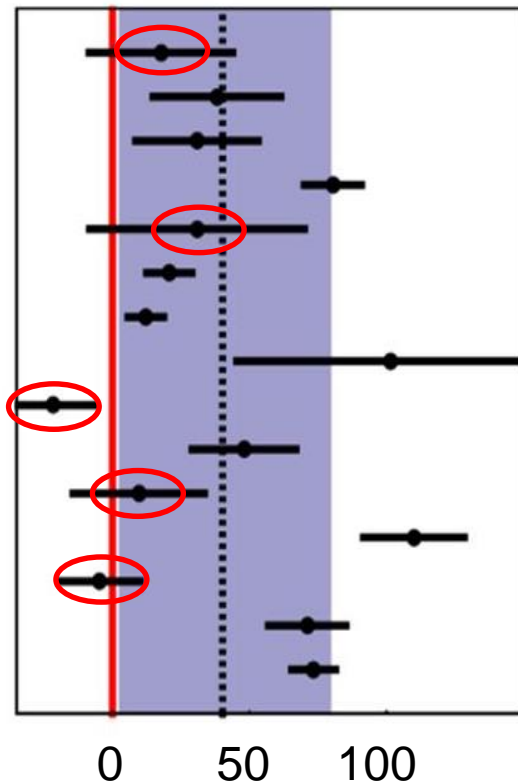
5 of 15 = accurate (coverage probability)



# Simulation of single-lab versus multi-lab studies

Effect of hypothermia on infarct volume in rodent models of stroke

1-lab studies

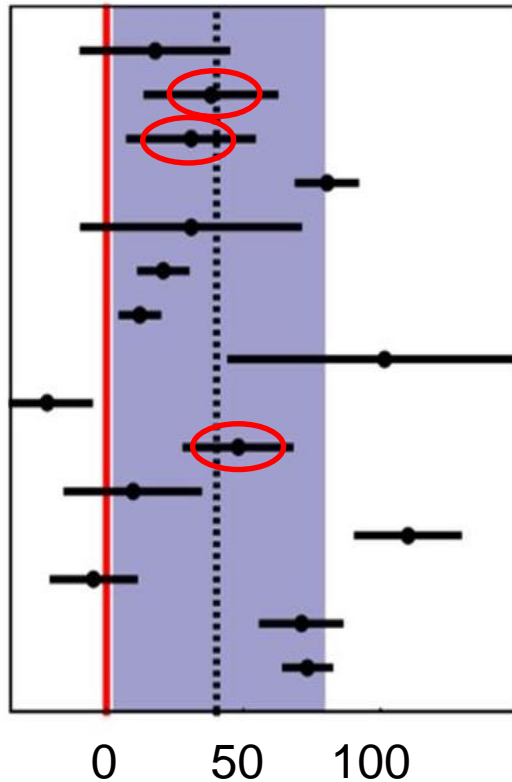


5 of 15 = false negatives

# Simulation of single-lab versus multi-lab studies

Effect of hypothermia on infarct volume in rodent models of stroke

1-lab studies

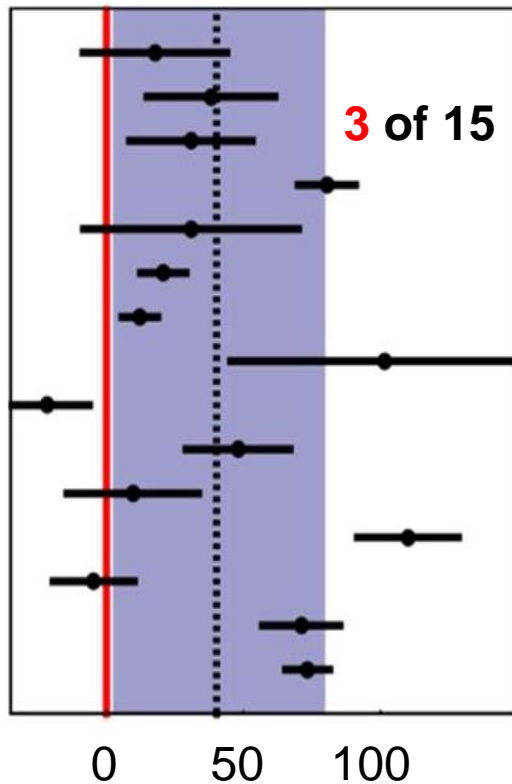


only **3** of 15 = significant and accurate

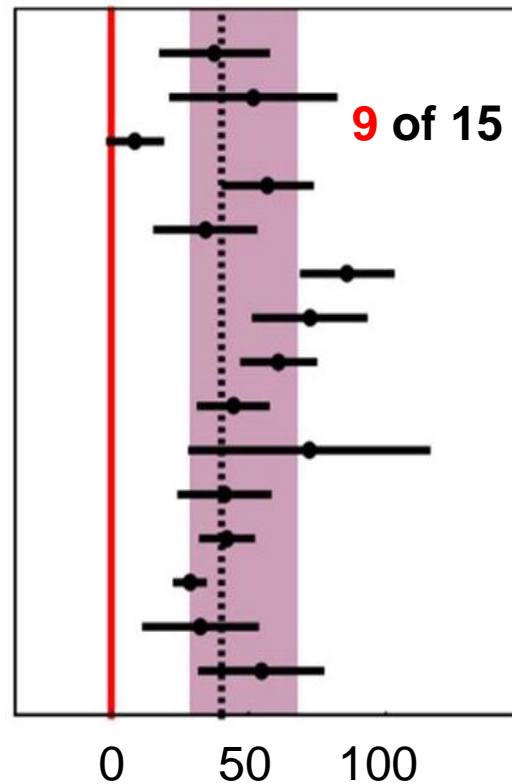
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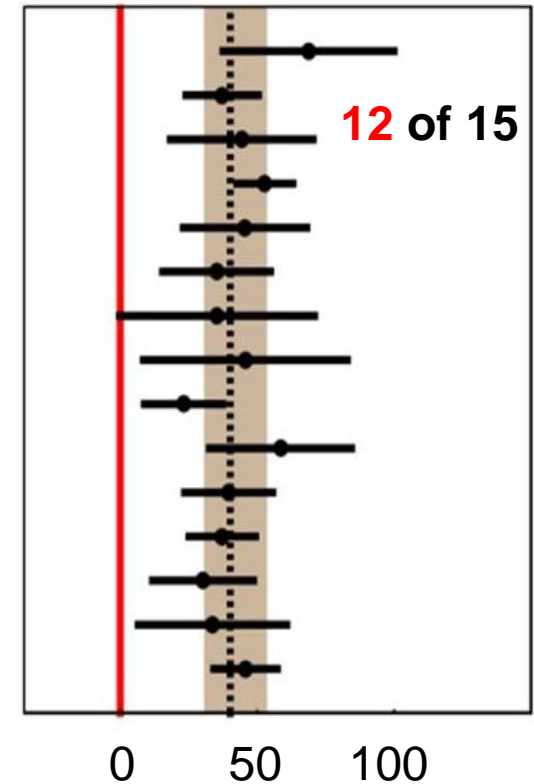
1-lab studies



2-lab studies

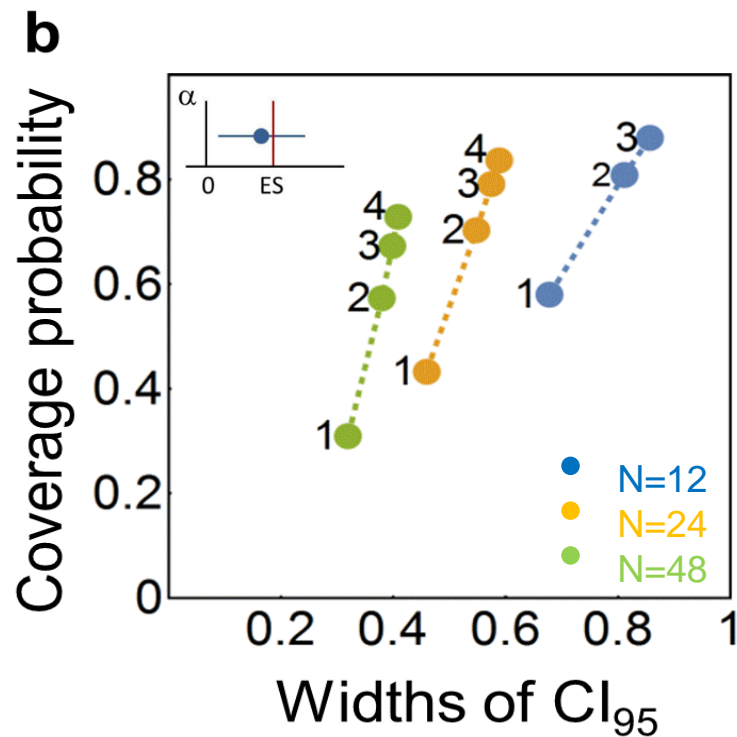


4-lab studies



# Simulation of single-lab versus multi-lab studies

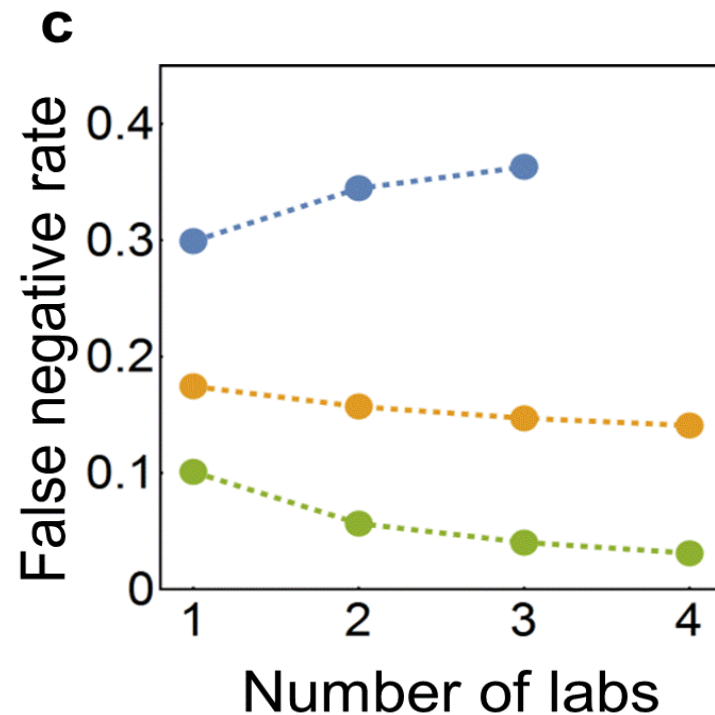
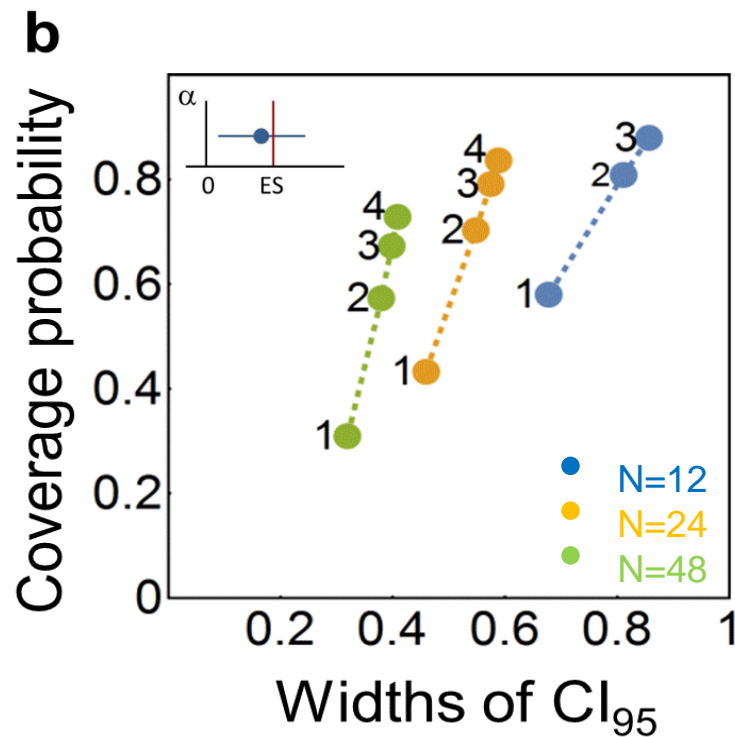
Coverage probability and false negative rate depending on sample size





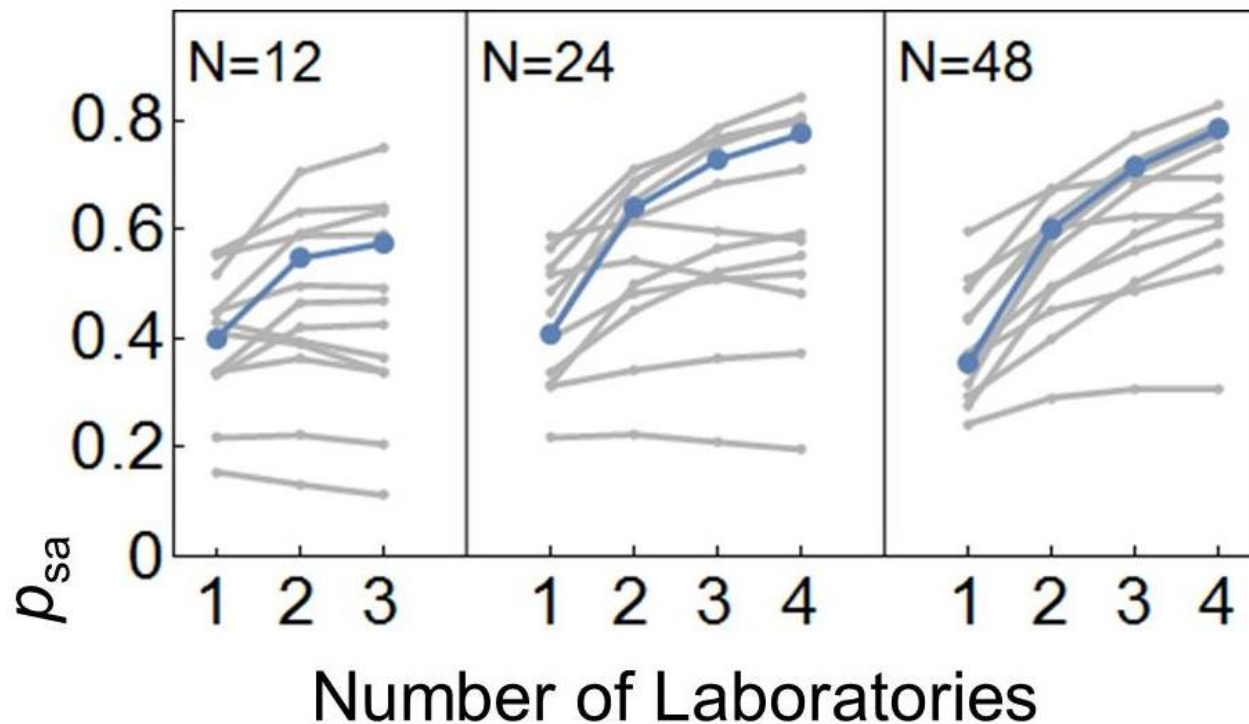
# Simulation of single-lab versus multi-lab studies

Coverage probability and false negative rate depending on sample size



# Simulation of single-lab versus multi-lab studies

Replications for 13 different interventions in animal models of stroke, myocardial infarction, and breast cancer



# Conclusions

- Reproducibility depends on the external validity of the results
- Standardized single-lab studies produce results of poor external validity
- Poor external validity leads to poor reproducibility

# Conclusions

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## Possible solutions:

- ▶ Adjusting p-values of single-lab studies by treatment x lab interaction term  
(Kafkafi *et al.* 2017 *Nat Methods*)
- ▶ Heterogenization of study samples in single-lab studies  
(Richter *et al.* 2010 *Nat Methods*)
- ▶ Multi-lab studies  
(Voelkl *et al.* 2018 *PLOS Biol*)



# Acknowledgments

## My lab

- Bernhard Voelkl (senior scientist)
- Lucile Vogt (PhD student)
- Helene Richter (former PhD student)

## Collaborators

- Emily Sena (Edinburgh)
- Yoav Benjamini (Tel Aviv)
- Joseph Garner (Stanford)

## Supported by



Schweizerische Eidgenossenschaft  
Confédération suisse  
Confederazione Svizzera  
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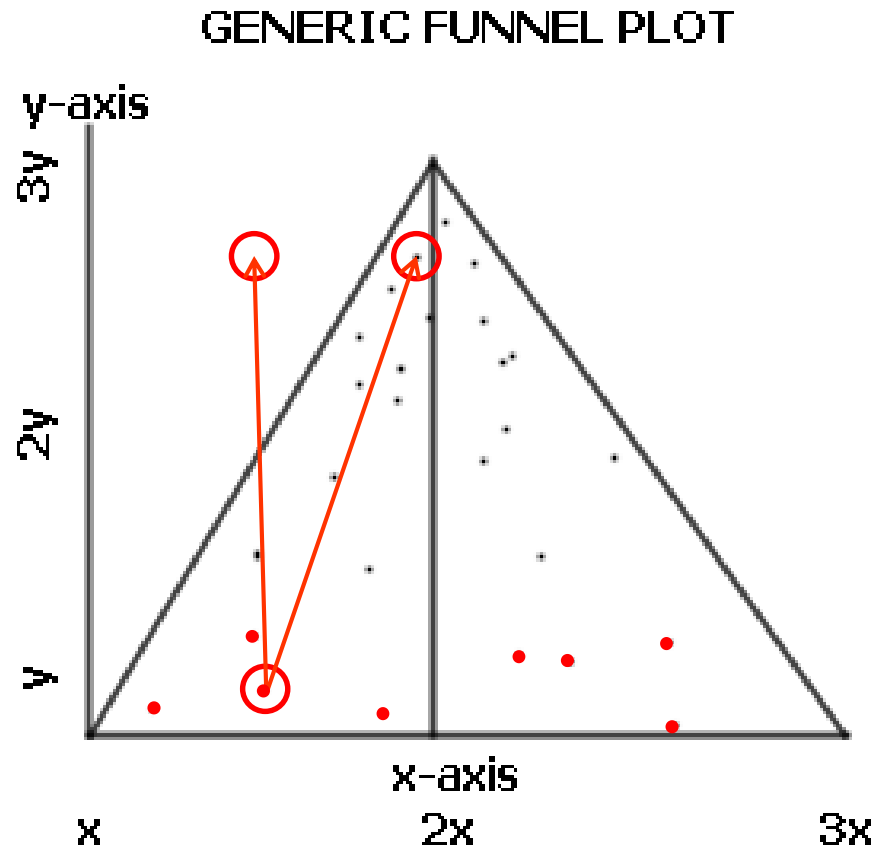


European Research Council



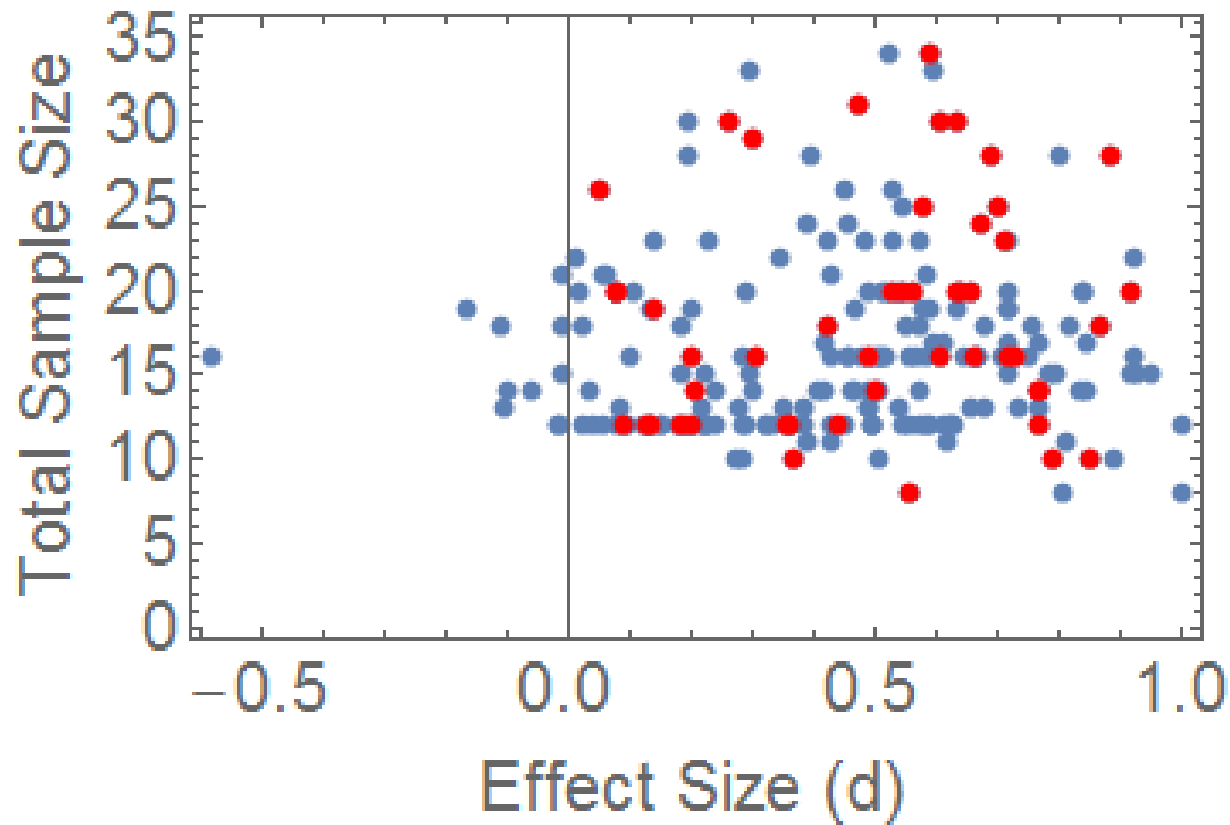
# Random variation or phenotypic plasticity?

Different predictions about effects of larger sample sizes



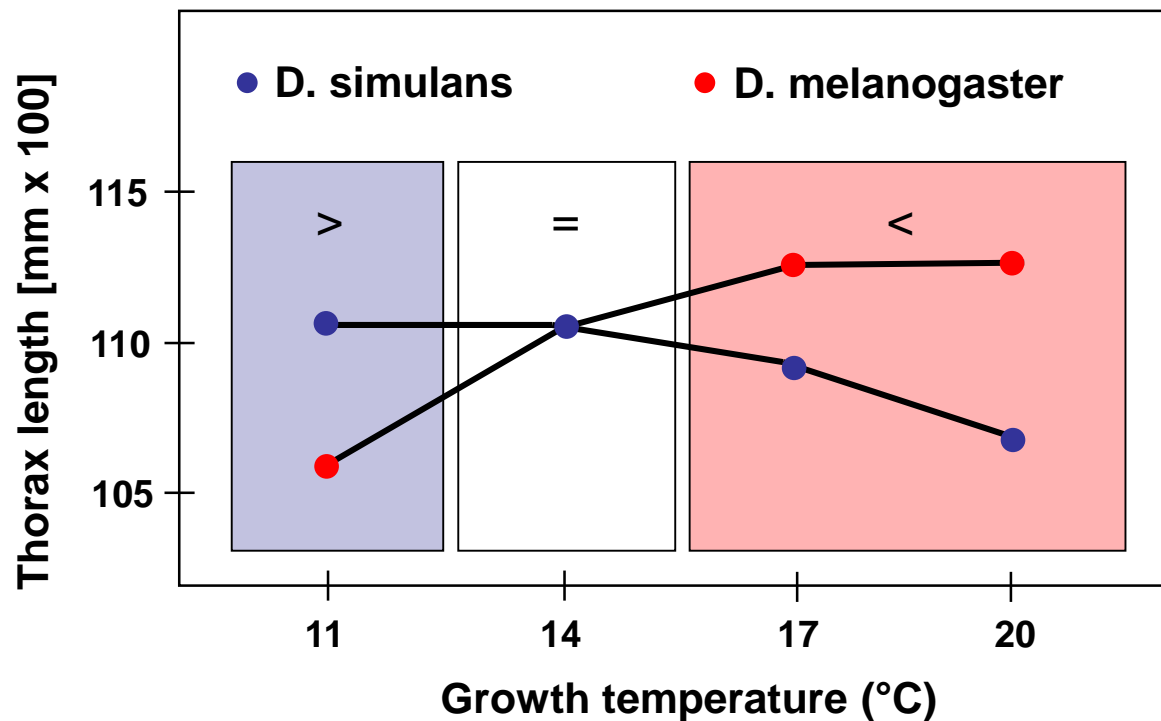
# Simulation of single-lab versus multi-lab studies

Coverage probability and false negative rate



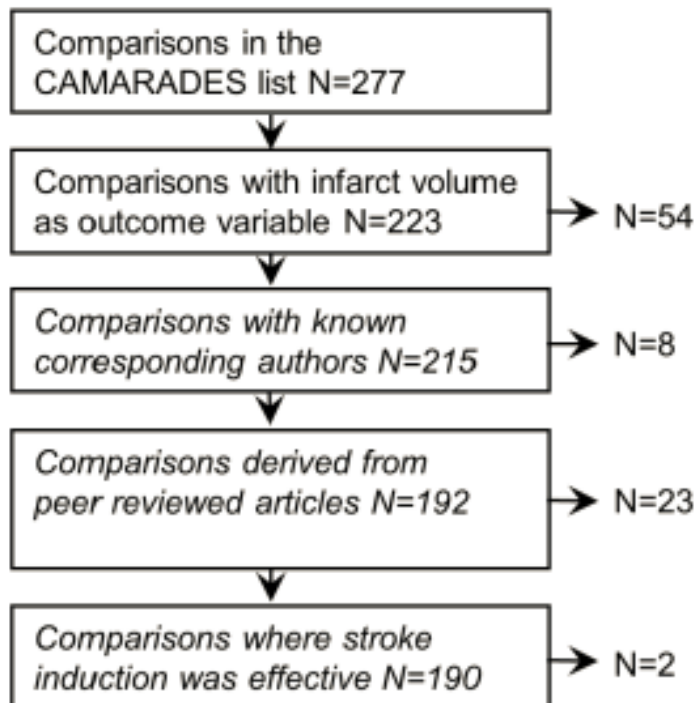
# Standardization and poor reproducibility

Phenotypic plasticity can induce treatment x environment interactions

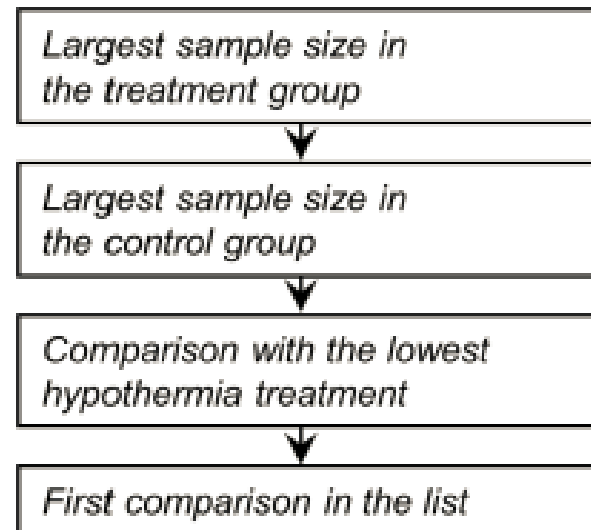


# Simulation of single-lab versus multi-lab studies

## Inclusion and exclusion criteria – hypothermia studies



*For selection of a single comparison per author:*



# Simulation of single-lab versus multi-lab studies

Further interventions (n=12) used to replicate hypothermia simulation

	Intervention	Outcome	Species	restricted	N
1	tPA	Infarct volume	Rat	Yes	57
2	Trastuzumab	Tumour volume ratio	Mouse	No	58
3	FK506	Infarct volume	Rat	Yes	31
4	Rosiglitazone 2	Infarct volume	Rodent	No	21
5	IL-1RA	Infarct volume	Rodent	No	37
6	Cardiosphere DC	EF (%)	Rodent	Yes	35
7	Estradiol	Infarct volume	Rat	Yes	24
8	Human MSC	Infarct volume	Rat	No	26
9	MK-801	Infarct volume	Rat	Yes	30
10	TMZ	Infarct volume	Rodent	No	26
11	c-kit CSC	EF (%)	Rodent	Yes	20
12	Rat BMSC	Infarct volume	Rat	No	25



# Simulation of single-lab versus multi-lab studies

## Results of random effects meta-analyses in R (*metafor* 1.9-9)

	Intervention	N	ES	S.E.	z	p	CL <sub>L</sub>	CL <sub>U</sub>	Q	p(Q)	log LH	dev	I <sup>2</sup>	H <sup>2</sup>
	Hypothermia	50	0.40	0.053	7.565	<0.0001	0.30	0.51	1801.0	<0.0001	-24.264	48.529	99.07	107.43
1	tPA	57	0.10	0.025	3.400	<0.0001	0.05	0.15	371.7	<0.0001	-13.026	26.052	91.36	11.57
2	Trastuzumab	58	0.24	0.031	7.915	<0.0001	0.18	0.31	3659.2	<0.0001	-28.569	57.137	99.02	101.73
3	FK 506	31	0.37	0.052	7.134	<0.0001	0.27	0.48	475.4	<0.0001	-6.010	12.020	98.98	97.64
4	Rosiglitazone 2	21	0.47	0.105	4.522	<0.0001	0.27	0.68	449.2	<0.0001	-14.091	28.182	98.55	68.97
5	IL-1RA	37	0.20	0.021	9.721	<0.0001	0.16	0.25	111.4	<0.0001	0.567	-1.133	62.91	2.70
6	Cardiosphere DC	35	-0.49	0.027	-18.15	<0.0001	-0.54	-0.44	278.4	<0.0001	11.992	-23.984	84.96	6.65
7	Estradiol	24	0.29	0.093	3.137	<0.0001	0.11	0.47	301.9	<0.0001	-19.168	38.337	99.57	234.07
8	Human MSC	26	0.24	0.058	4.102	<0.0001	0.12	0.35	844.3	<0.0001	-6.039	12.079	99.53	215.03
9	MK 801	30	0.27	0.050	5.431	<0.0001	0.18	0.37	624.2	<0.0001	-4.991	9.983	99.99	9074.30
10	TMZ	26	0.31	0.121	2.545	0.0109	0.07	0.55	1365.0	<0.0001	-28.487	56.974	100.00	355403
11	c-kit CSC	20	-0.33	0.032	-10.36	<0.0001	-0.39	-0.27	43.5	0.0011	9.671	-19.342	48.50	1.94
12	Rat BMSC	25	0.29	0.134	2.045	0.0409	0.01	0.56	1434.7	<0.0001	-25.446	50.892	99.51	202.11